

HEK293T | 300189

Thawing and Culturing Cells

1. Thaw the vial quickly in a 37°C water bath, and transfer the cells to a pre-warmed T25 flask containing 5-10 ml of DMEM supplemented with 10% FBS.
2. Incubate the cells for 24-48 hours at 37°C in 5% CO₂ to allow the cells to recover and reach confluence.
3. Once the cells are confluent, replace the medium with DMEM supplemented with 10% FBS.
4. After 24-48 hours, the cells should be ready for passaging. Remove the FBS and replace with DMEM supplemented with 10% FBS.
5. Seed the cells into a new T25 flask containing 5-10 ml of DMEM supplemented with 10% FBS.
6. Incubate the cells for 24-48 hours at 37°C in 5% CO₂ to allow the cells to recover and reach confluence.
7. Once the cells are confluent, replace the medium with DMEM supplemented with 10% FBS.
8. After 24-48 hours, the cells should be ready for passaging. Remove the FBS and replace with DMEM supplemented with 10% FBS.

Incubation Atmosphere 37°C, 5% CO₂, humidified air

Flask Coating None

Freezing Procedure Harvest cells into a T25 flask, wash with PBS, and resuspend in freezing medium. Aliquot into 1 ml vials and store at -80°C.

Shipping Conditions Cells should be shipped at -80°C in dry ice.

Storage Conditions Cells should be stored at -80°C in liquid nitrogen.

Genotype / Phenotype / HLA

Sterility Cells are tested for mycoplasma contamination using PCR. Cells are also tested for endotoxin contamination.