

Product sheet

Hep-CLS-1H | 400197

Culture Medium Ham's F12, w: 1.0 mM β -mercaptoethanol, w: 1.0 mM $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, w: 1.1 g/L NaHCO_3 (Cytion 820600a)

Supplements β -mercaptoethanol 10% FBS

Dissociation Reagent β -mercaptoethanol

Subculturing Cells are cultured in Ham's F12 medium supplemented with 10% FBS and 100 U/ml penicillin, 100 U/ml streptomycin, and 100 U/ml nystatin. For subculturing, cells are trypsinized with 0.25% trypsin-EDTA (Cytion 820600a) for 5 minutes at 37°C. Cells are then washed with PBS and resuspended in fresh medium.

Fluid renewal 3-5 times

Freeze medium β -mercaptoethanol, β -mercaptoethanol, β -mercaptoethanol (10% FBS) + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the cells rapidly in a 37°C water bath. Transfer the cells to a pre-warmed medium.
 2. Allow the cells to settle at the bottom of the tube. Centrifuge at 300 x g for 5 minutes.
 3. Wash the cells with PBS. Resuspend in fresh medium.
 4. Seed the cells into a pre-warmed flask. Add medium to 70% confluence.
 5. Incubate the cells at 37°C in 5% CO_2 atmosphere.
 6. Monitor the cells for confluence. Refresh the medium when needed.
 7. Harvest the cells when they reach 80-90% confluence.
 8. Perform a final wash with PBS before harvesting.

Incubation Atmosphere 37°C, 5% CO_2

Flask Coating β -mercaptoethanol, β -mercaptoethanol

