

Product sheet

**HEP-56.1C | 400203**

**HEP-56.1C | 400203**

**Description** HEP-56.1C is a cell line derived from a human liver carcinoma, established in 1965. It is a continuous cell line that grows in suspension culture. The cells are of epithelial origin and are characterized by their ability to form spheroids in suspension culture. HEP-56.1C is a cell line derived from a human liver carcinoma, established in 1965. It is a continuous cell line that grows in suspension culture. The cells are of epithelial origin and are characterized by their ability to form spheroids in suspension culture. HEP-56.1C is a cell line derived from a human liver carcinoma, established in 1965. It is a continuous cell line that grows in suspension culture. The cells are of epithelial origin and are characterized by their ability to form spheroids in suspension culture.

**Organism** HEP-56.1C

**Tissue** Liver

**Disease** Hepatocellular carcinoma

**Synonyms** HEP-56.1C, 56.1C, 56.1c

**HEP-56.1C | 400203**

**Breed/Subspecies** C57BL/6J

**Age** 1-3 months

**Gender** Male

**Morphology** Epithelial cells

**Growth properties** Suspension culture

**HEP-56.1C | 400203**

**Citation** HEP-56.1C (HEP-56.1C) Cytion 400203

**Biosafety level** 1

**NCBI\_TaxID** 10090

**CellosaurusAccession** CVCL\_5768

**HEP-56.1C | 400203**

# Hep-56.1C | 400203

## Characteristics

**Culture Medium** DMEM, w: 4.5 g/L D-glucose, w: 4 mM L-glutamine, w: 3.7 g/L NaHCO<sub>3</sub>, w: 1.0 mM β-mercaptoethanol (Cytion 820300a)

**Supplements** 10% FBS

**Dissociation Reagent** Trypsin

**Subculturing** Seed cells into T25 flasks with 3 ml of DMEM + 10% FBS. When cells reach 70-80% confluency, trypsinize and seed into 37°C incubator.

**Seeding density** 1 x 10<sup>4</sup> cells/cm<sup>2</sup>

**Fluid renewal** 3-5 days

**Post-Thaw Recovery** After thawing, seed cells into T25 flasks with 3 ml of DMEM + 10% FBS. Allow cells to recover for 24 hours before use.

**Freeze medium** DMEM + 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw vials quickly in a 37°C water bath.
  2. Dilute cells into 10 ml of DMEM + 10% FBS.
  3. Seed cells into T25 flasks with 3 ml of medium.
  4. Allow cells to recover for 24 hours.
  5. After 24 hours, replace medium with fresh DMEM + 10% FBS.
  6. When cells reach 70-80% confluency, trypsinize and seed into new flasks.
  7. Seed cells into T25 flasks with 3 ml of DMEM + 10% FBS.
  8. Allow cells to recover for 24 hours.

