

Product sheet

**XXXXX S-117 | 300329**

**XXXXX XXXXX**

<b>Description</b>	XXXX XXXXXX XXXXXXXX XXXXXXXX XXXXXXX XXXXX XX XXXX XX 47.
<b>Organism</b>	XXX
<b>Tissue</b>	XXXXXXXXXX
<b>Disease</b>	XXXXXXXX
<b>Synonyms</b>	S-117, S117

**XXXXXXXXXXXX**

<b>Age</b>	47 XXXX
<b>Gender</b>	XXXX
<b>Morphology</b>	XXXX XXXXXXXXXXXXX, XXXXX XXXXXXXXXXXXX
<b>Growth properties</b>	XXX

**XXXXXXXXX XXXXXXXXXXXXXXX**

<b>Citation</b>	S-117 (XXXX XXXXXXX Cytion 300329)
<b>Biosafety level</b>	1
<b>NCBI_TaxID</b>	9606
<b>CellosaurusAccession</b>	CVCL_1674

**XXXXXXXXX XXX-XXXXXXXXXXXX**

<b>Tumorigenic</b>	XX, XXXXXXXX XXXXXXXX
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**XXXXXXXX**

Product sheet

**HEK293T-S-117 | 300329**

**Culture Medium** RPMI 1640, w: 2.0 mM  $\beta$ -mercaptoethanol, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion  $\beta$ -mercaptoethanol Cytion 820700a)

**Supplements**  $\beta$ -mercaptoethanol 10% FBS

**Dissociation Reagent**  $\beta$ -mercaptoethanol

**Subculturing** Cells are cultured in 25 cm<sup>2</sup> flasks in RPMI 1640 medium supplemented with 10% FBS and 2.0 mM  $\beta$ -mercaptoethanol. Cells are passaged when they reach 80-90% confluency. For subculturing, cells are trypsinized and resuspended in 10 ml of RPMI 1640 medium supplemented with 10% FBS and 2.0 mM  $\beta$ -mercaptoethanol. Cells are then seeded into new flasks at a density of 1 x 10<sup>4</sup> cells per flask.

**Split ratio** 1:4 or 1:8

**Seeding density** 1 x 10<sup>4</sup> cells per flask

**Fluid renewal** 2-3 times per week

**Post-Thaw Recovery** After thawing, cells are seeded into a 25 cm<sup>2</sup> flask in RPMI 1640 medium supplemented with 10% FBS and 2.0 mM  $\beta$ -mercaptoethanol. Cells are allowed to recover for 24 hours before being passaged.

**Freeze medium** RPMI 1640 medium supplemented with 10% FBS and 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the vial in a 37°C water bath.
  2. Add 10 ml of RPMI 1640 medium supplemented with 10% FBS and 2.0 mM  $\beta$ -mercaptoethanol to the vial.
  3. Gently mix the cells and seed them into a 25 cm<sup>2</sup> flask.
  4. Allow the cells to recover for 24 hours.
  5. Pass the cells into a new flask when they reach 80-90% confluency.
  6. Seed cells at a density of 1 x 10<sup>4</sup> cells per flask.
  7. Maintain cells in RPMI 1640 medium supplemented with 10% FBS and 2.0 mM  $\beta$ -mercaptoethanol.
  8. Pass cells when they reach 80-90% confluency.



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**A\***: 01:01:01  
**B\***: '37:01:01  
**C\***: 06:02:01  
**DRB1\***: 11:01:01  
**DQA1\***: 05:05:01  
**DQB1\***: 03:01:01  
**DPB1\***: 04:01:01  
**E**: 01:01:01