

XXXXHROC60 | 300827

Tumorigenic XX, XXXXXXXX XXXXXXXX XX XXXXX XXXXXXXX

Viruses XXX XXXXXXXX XXXXXXXX XXXX XXX SV40, JC/BK, HBV, HCV, HIV.

Ploidy status XXXXXXXXXX

MSI-status MSS

Mutational profile XXXXXX KRAS

XXXXXX

Culture Medium DMEM:Ham's F12 (1:1), w: 3.1 g/L XXXXXXX, w: 2.5 mM L-XXXXXXX, w: 15 mM HEPES, w: 0.5 mM XXXX XXXXXXX, w: 1.2 g/L NaHCO3 820400a)

Supplements XXXX XXXXX 10% FBS

Dissociation Reagent XXXXXXX

Doubling time 29 XXXXX

Subculturing XXXX XX XXXXXXX XXXX XXXXXXX XXXXXXX XXXXX XXXX 4-PBS XXX XXXX XXXXXXX XXXX XXXXXXX T25, XXXXX 4-3-5 4' 4-PBS, XXXXX XXXX 3 XXXXX. XXXX XX XXXXXXX XXXXXXX, XXXX XX XXXXXXX XXXXXXX XXXXXXX XXX XXXXXXX XXXX XXXXXXX XXXXXXX XXXX XXXX XXXXXXX XXXXXXX XXXX

Seeding density 2×10^4 ^{10⁶} XXXXX/XX

Fluid renewal XXX 3 XXX 5 XXXXX

Post-Thaw Recovery 1 XXX 2 XXXXXXX

Freeze medium XXXXXXX XXXXXXX XXXXXXX, XXXX XXXXXXX XXXXXXX XXXXXXX XXXX (XXXXX FBS) + 10% DMSO XXXX XXXXXXX XXXXXXX XXXXXXX XXXXXXX XXXXXXX, XXXX

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Thawing and Culturing Cells

1. Thaw the vial quickly in a 37°C water bath. Transfer the cells to a pre-warmed T75 flask containing 10 ml of complete DMEM medium. Centrifuge at 300 x g for 3 minutes. Remove the medium and replace with 10 ml of fresh complete DMEM medium.
2. Incubate the cells at 37°C in 5% CO₂ until they reach 70-80% confluency. The cells should be ready for passaging after 2-3 days.
3. For passaging, trypsinize the cells and seed them into a new T75 flask with 10 ml of complete DMEM medium. The cells should reach 70-80% confluency again within 2-3 days.
4. For freezing, trypsinize the cells and resuspend them in 1 ml of freezing medium. Seed into a 1.5 ml microcentrifuge tube and freeze at -80°C.
5. For thawing, thaw the vial quickly in a 37°C water bath. Transfer the cells to a pre-warmed T75 flask containing 10 ml of complete DMEM medium. Centrifuge at 300 x g for 3 minutes. Remove the medium and replace with 10 ml of fresh complete DMEM medium.
6. Incubate the cells at 37°C in 5% CO₂ until they reach 70-80% confluency. The cells should be ready for passaging after 2-3 days.
7. For passaging, trypsinize the cells and seed them into a new T75 flask with 10 ml of complete DMEM medium. The cells should reach 70-80% confluency again within 2-3 days.
8. For freezing, trypsinize the cells and resuspend them in 1 ml of freezing medium. Seed into a 1.5 ml microcentrifuge tube and freeze at -80°C.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating None

Freezing Procedure Resuspend cells in 1 ml of freezing medium, seed into a 1.5 ml microcentrifuge tube, and freeze at -80°C.

Shipping Conditions -80°C

Storage Conditions -150°C, 196 liquid nitrogen

HEK293T / HEK293T / HLA

Sterility The cells are provided in a sterile, cryoprotected medium. They are not tested for mycoplasma contamination. PCR genotyping is available upon request.