

Product sheet

**XXXX B-LCL-CDG5 | 302016**

**XXXX XXXX**

**Description** B-LCL-CDG5 XXXX XX XXXX XX XXXXXXXXXXXXXXX B XXXXX XXXXX XX XXX EBV, XXXXXXX XXXXXXX XX PMM2-CDG, XXXXX XXXXX XXXXXXXXXXXXXXX  
XXXX XXXX B XXXXXXX XX XXX EBV, B-LCL-CDG5 XXXX XXXX XXXXX XXXX XXXXXXXXXXX XXXXXXXXXXXXXXX XXXXXXXXXXXXXXX \*PMM2\* .

**Organism** XXXX

**Tissue** XX XXXXX

**Disease** XXXXX

**Applications** XXXXXXX XX XXXXXXX CDG XXXX XXXXX, XXXXXXX XXXXXXX (XXXX, XXXXXXX XX XXX XXX B), XXXXXXX XX XXXXXXX XXXXXXXXXXXXXXX XXXXX

**XXXXXXXXXX**

**Gender** XXXXX

**Ethnicity** XXXXXXX

**Morphology** XXXX XXXXXXX

**Cell type** XXXXXXXXX B

**Growth properties** XXXX, XXXXX

**XXXXXXXXX XXXXXXXXXXXXXXX**

**Citation** B-LCL-CDG5 (XXXX XXXXXXX Cytion 302016)

**Biosafety level** 2

**NCBI\_TaxID** 9606

**XXXXXXXXX XXX- XXXXXXXXXXXXXXX**

**Viruses** XXXXXXXXXXXX EBV

### HEK293T-B-LCL-CDG5 | 302016

#### HEK293T

**Culture Medium** RPMI 1640, w: 2.0 mM  $\beta$ -mercaptoethanol, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion 820700a)

**Supplements** 10% FBS

**Subculturing** 1:5

**Fluid renewal**

**Post-Thaw Recovery**

**Freeze medium** 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells rapidly in a 37°C water bath.
  2. Centrifuge cells at 300 x g for 3 minutes.
  3. Resuspend cells in 15 ml of fresh culture medium.
  4. Seed cells into a 75 cm<sup>2</sup> flask at 70% confluency.
  5. Incubate cells at 37°C in 5% CO<sub>2</sub>.
  6. Harvest cells when they reach 80-90% confluency.
  7. Wash cells with PBS.
  8. Harvest cells using trypsin.

**Incubation Atmosphere** 37°C, 5% CO<sub>2</sub>

**Flask Coating**

