

Product sheet

XXXX B-LCL-CDG3 | 302014

XXXX XXXX

Description B-LCL-CDG3 XXXX XX XXXX XX XXXXXXXXXXXXXXX B XXXXX XXXXX XX XXX EBV, XXXXXXX XXXXXXX XX PMM2-CDG, XXXXX XXXXX XXXXXXXXXXXXXXXXXXXXXXX
XXXX XXXX B XXXXXXX XX XXX EBV, B-LCL-CDG3 XXXX XXXXX XXXX XXXXXXX XXXX XXXXXXXXXXXXXXXXXXXXXXX XX XXXXXXX *PMM2*. XX XXXX

Organism XXX

Tissue XX XXXXX

Disease XXXXXXX XXXXXXX XXXXXXXXXXXXXXX

Applications XXXXXXX XX XXXXXXX CDG XXXX XXXXX, XXXXXXX XXXXXXX (XXXX, XXXXXXX XX XXX XX B), XXXXXXX XX XXXXXXX XXXXXXXXXXXXXXX XXXXX

XXXXXXXXXX

Gender XXXX

Ethnicity XXXXXXX

Morphology XXXX XXXXXXX

Cell type XXXXXXX B

Growth properties XXXX, XXXXX

XXXXXXXXX XXXXXXXXXXXXXXX

Citation B-LCL-CDG3 (XXXX XXXXXXX Cytion 302014)

Biosafety level 2

NCBI_TaxID 9606

XXXXXXXXX XXX- XXXXXXXXXXXXXXX

Viruses XXXXXXXXXXXX EBV

HEK293T-B-LCL-CDG3 | 302014

HEK293T

Culture Medium RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a)

Supplements 10% FBS

Subculturing 5:5:5 ratio

Fluid renewal 3x per week

Post-Thaw Recovery 24h

Freeze medium 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw cells rapidly in a 37°C water bath.
 2. Centrifuge cells at 300 x g for 3 minutes.
 3. Resuspend cells in 15 ml of fresh medium.
 4. Seed cells into a T25 flask at 70% confluency.
 5. Incubate cells at 37°C in 5% CO₂.
 6. Harvest cells when they reach 80-90% confluency.
 7. Wash cells with PBS.
 8. Harvest cells using trypsin.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating Poly-D-Lysine

