

Product sheet

XXXX OVCAR-8 | 305383

XXXX XXXX

Description OVCAR-8 XXXX XX XXXX XX XXXXXXXXXXXX XXXXXX XXXXXXXX XXXXXX XXXXXX XX XXXXXXXXXXXXXXXXXXXX XXXXXX XXXXX XXXXXXX. XX XXXXX XX XXXXX XXXXXXX XXXXXXXXXXXX
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OVCAR-8 XXXX XXXX XXXXX XXXXXXX XXX- XXXXXXX XXXXXXX XXXXXXXXXXXXXXXXXXXX, XXXXXXX XXXXXXXXXXX XXXXXXXXXXX XXXXX XXXXXXXXXXXXXXX XX XXXXXXXXXXX XXXXXXXXXXXXXXX

Organism XXX

Tissue XXXXX

Disease XXXXXXXXXXXXXXX XX XXXXXXX

Synonyms OVCAR 8, NIH:OVCAR-8, OVCAR8, OvcAR8, OVCAR.8, OVCA8, OVCAR-8/EGFP_LC3

XXXXXXXXXXXX

Age 64 XXXXX

Gender XXXXX

Ethnicity XXXXXXX

Morphology XXXXX XXXXXXX

Growth properties XXXX

XXXXXXXXXX XXXXXXXXXXXXXXXXXXXX

Citation OVCAR-8 (XXXX XXXXXXX Cytion 305383)

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_1629

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Mutational profile CTNNB1, p.Gln26Arg (c.77A>G), ERBB2, p.Gly776Val (c.2327G>T), c.376-1G>A (p.Tyr126_Lys132del, c.376_396del21),

Culture Medium RPMI 1640, w: 2.1 mM NaHCO3, w: 2.0 g/L NaHCO3 (Cytion 820700a)

Supplements 10% FBS

Dissociation Reagent

Doubling time 24-32

Split ratio 1:4 1:4

Seeding density $3-4 \times 10^4$ cells/cm²

Freeze medium (FBS) + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a 37°C water bath, then transfer to a 15 mL centrifuge tube.
2. Centrifuge at 300 x g for 3 minutes, remove supernatant, and resuspend in 10 mL of culture medium.
3. Seed cells into a 75 cm² tissue culture flask at a density of 15 x 10⁴ cells/cm².
4. Incubate cells in a humidified 5% CO₂ incubator at 37°C.
5. Monitor cell growth and confluency.
6. Once cells reach 70-80% confluency, passage them into a new flask.
7. Repeat the process for subsequent passages.
8. Maintain cells in a humidified 5% CO₂ incubator at 37°C.

