

Product sheet

KU812 | 305306

General Information

Description	<p>KU812 is a cell line derived from a patient with Chronic Myeloid Leukemia (CML) who was treated with Imatinib. The cell line is characterized by the presence of the BCR-ABL1 fusion gene. It is a Philadelphia chromosome positive (Ph+) cell line. The cell line is maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin (PS). The cell line is a continuous cell line and is used for research purposes.</p> <p>KU812 is a cell line derived from a patient with Chronic Myeloid Leukemia (CML) who was treated with Imatinib. The cell line is characterized by the presence of the BCR-ABL1 fusion gene. It is a Philadelphia chromosome positive (Ph+) cell line. The cell line is maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin (PS). The cell line is a continuous cell line and is used for research purposes.</p>
Organism	Human
Tissue	Leukemia
Disease	Chronic Myeloid Leukemia (CML), BCR-ABL1
Synonyms	Ku812, KU-812, KU.812, KU 812

Cell Line Characteristics

Age	38 years
Gender	Male
Ethnicity	White
Morphology	Granulocytic
Cell type	Leukemia
Growth properties	Adherent

References and Safety

Citation	KU812 (ATCC CCL-222) Cytion 305306
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0379

HEK293T KU812 | 305306

HEK293T KU812 - HEK293T KU812

Antigen expression CD3, ANPEP (CD13)

Mutational profile TP53, p.Lys132Arg (c.395A>G), BCR-ABL, 14 BCR 2 ABL1 (TP53)

Karyotype Ph1 (Ph1)

HEK293T

Culture Medium RPMI 1640, w: 2.0 mM NaH_2PO_4 , w: 2.0 g/L NaHCO_3 (Cytion 820700a)

Supplements 10% FBS, 2.5 $\mu\text{g}/\text{ml}$ HEPES

Subculturing 15 μl PBS (3-5 μl)

Seeding density 3×10^5 cells/ μl

Fluid renewal 2-3 times

Freeze medium (FBS) + 10% DMSO

Cell Culture KU812 | 305306

Thawing and Culturing Cells

1. **Thawing:** Thaw the vial rapidly in a 37°C water bath. Transfer the cells to a pre-warmed medium.
2. **Centrifugation:** Centrifuge the cells at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of pre-warmed medium.
3. **Seeding:** Seed the cells into a T25 flask containing 10 ml of pre-warmed medium.
4. **Incubation:** Incubate the cells at 37°C in 5% CO₂ until they reach 70% confluency.
5. **Passaging:** When cells reach 70% confluency, passage them into a new T25 flask.
6. **Media Change:** Change the medium every 2-3 days.
7. **Freezing:** For long-term storage, freeze the cells in a freezing medium.
8. **Quality Control:** Perform regular quality control checks to ensure cell viability and identity.

Incubation Atmosphere

37°C, 5% CO₂, humidified

Flask Coating

Flasks should be coated with a cell culture adhesive (e.g., Cell Culture Adhesive) before use.

Freezing Procedure

For long-term storage, freeze the cells in a freezing medium at -80°C.

Shipping Conditions

Shipping conditions: -78°C

Storage Conditions

Storage conditions: -150 to 196 K

Cell Culture / **Cell Culture** / **HLA**

Sterility

Cells are provided in a sterile, cryoprotective medium.

Cells are provided in a sterile, cryoprotective medium.