





**HEK293T HCC1143 | 305545**

**Thawing and Culturing Cells**

1. **Thawing:** Thaw the vial containing the cells in a 37°C water bath. Transfer the cells to a pre-warmed T25 flask containing 10 ml of complete DMEM medium.
2. **Seeding:** Seed the cells into a T25 flask containing 10 ml of complete DMEM medium. The cell density should be approximately 1.5 x 10<sup>5</sup> cells per flask.
3. **Incubation:** Incubate the cells in a humidified 5% CO<sub>2</sub> incubator at 37°C. The cells should reach 70-80% confluency within 2-3 days.
4. **Passaging:** Once the cells reach 70-80% confluency, they can be passaged into a new T25 flask. Use trypsin-EDTA to detach the cells.
5. **Media Change:** Change the medium to fresh complete DMEM medium. The volume of medium should be 10 ml per T25 flask.
6. **Subculturing:** Subculture the cells into a new T25 flask. The cell density should be approximately 1.5 x 10<sup>5</sup> cells per flask.
7. **Media Change:** Change the medium to fresh complete DMEM medium. The volume of medium should be 10 ml per T25 flask.
8. **Storage:** The cells can be stored in a liquid nitrogen vapor phase for long-term storage. The storage medium should be complete DMEM medium with 10% FBS.

**Incubation Atmosphere**

37°C, 5% CO<sub>2</sub>, humidified

**Shipping Conditions**

Cells should be shipped in a dry ice container at -78°C.

**Storage Conditions**

Cells should be stored in a liquid nitrogen vapor phase at -150°C. The storage medium should be complete DMEM medium with 10% FBS.

**HEK293T / HEK293T / HLA**

**Sterility**

Cells are provided in a sterile, sealed vial. The medium is sterile and contains no antibiotics.

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