

Product sheet

HEK293T P388 | 305226

Culture Medium RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a)

Supplements 10% FBS

Subculturing Seed cells into fresh medium at 70-80% confluency. Use 1:2 to 1:10 split ratios. Wash cells with PBS before trypsinization. Add trypsin to cells for 2-5 minutes at 37°C. Stop reaction with 10% FBS medium. Resuspend cells in fresh medium.

Freeze medium RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a), 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw vials quickly in a 37°C water bath. Transfer cells to a pre-warmed medium.
 2. Centrifuge cells at 300 x g for 3 minutes. Resuspend cells in fresh medium.
 3. Seed cells into a 15 cm² flask at 10⁶ cells per flask. Incubate at 37°C.
 4. Monitor cell growth. Split cells at 70-80% confluency.
 5. Use 1:2 to 1:10 split ratios. Wash cells with PBS before trypsinization.
 6. Add trypsin to cells for 2-5 minutes at 37°C. Stop reaction with 10% FBS medium.
 7. Resuspend cells in fresh medium. Seed cells into a new flask.
 8. Repeat the process for subsequent passages.

Incubation Atmosphere 37°C, 5% CO₂, humidified

Flask Coating Adherent cells: none. Non-adherent cells: none.

Freezing Procedure Harvest cells at 70-80% confluency. Wash cells with PBS. Add trypsin to cells for 2-5 minutes at 37°C. Stop reaction with 10% FBS medium. Resuspend cells in freeze medium.

Shipping Conditions Store cells at -78°C. Ship cells in a dry ice container.

