

HCT-15 Cells | 300229

General information

Description

HCT-15 cells are derived from the adenocarcinoma of the colon of a 44-year-old Caucasian male. This cell line, developed in the early 1970s, is widely utilized in the field of cancer research, especially for exploring the biology and treatment of colorectal cancer.

Morphologically, HCT-15 cells are characterized by an epithelial-like appearance with a tendency to grow both as a monolayer and in clusters, displaying significant cellular heterogeneity. This feature mirrors the varied cellular environments found in solid tumors, making HCT-15 a valuable model for studying tumor dynamics and cellular interactions within the tumor microenvironment.

Genotypically, HCT-15 cells exhibit a hyperdiploid karyotype with multiple chromosomal aberrations, typical of many colorectal cancers. These include mutations in key oncogenes and tumor suppressor genes, such as mutations in the KRAS gene and deletions affecting the p53 pathway, which are implicated in the pathogenesis and progression of colorectal cancer. These genetic traits make HCT-15 cells a crucial tool for investigating genetic and molecular mechanisms associated with cancer progression, metastasis, and resistance to therapies.

The broad use of HCT-15 cells in research has led to significant insights into the molecular pathways involved in colorectal cancer, enhancing our understanding of disease mechanisms and aiding in the development of targeted therapies.

Organism Human

Tissue Colorectal

Disease Adenocarcinoma

Synonyms HCT 15, HCT.15, HCT15

Characteristics

Age 67 years

Gender Male

Morphology Epithelial-like

Growth properties Adherent

Regulatory Data

Citation HCT-15 (Cytion catalog number 300229)

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Biosafety level 1**NCBI_TaxID** 9606**CellosaurusAccession** CVCL_0292**Biomolecular Data****Antigen expression** The cells are positive for keratin by immunoperoxidase staining.**Tumorigenic** In nude mice**Viruses** Reverse Transcriptase negative**Handling****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS**Dissociation Reagent** Accutase**Doubling time** 15 hours**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Seeding density** 1 to 2 x 10⁴ cells/cm²**Fluid renewal** 2 to 3 times per week**Post-Thaw Recovery** Fast

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Freeze medium

As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Flask Coating

None

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

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Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.