

**D341Med Cells | 305136**

**General information**

**Description**

The D341 Med cell line was established in 1988 by Friedman et al. from tumor tissue extracted from a 3-year-old boy diagnosed with medulloblastoma. Medulloblastoma is a highly malignant pediatric brain tumor predominantly occurring in the cerebellum. This cell line is crucial for research due to its origin from a common type of childhood brain cancer, providing insights into the tumor biology and genetics specific to pediatric cases. D341 Med has been extensively utilized in studies aimed at understanding the molecular and cellular mechanisms of medulloblastoma, including investigations into the genetic mutations and signaling pathways that contribute to tumorigenesis and treatment resistance.

In addition to its role in basic research, the D341 Med cell line has been instrumental in preclinical studies assessing new therapeutic approaches for medulloblastoma. Its genetic profile, which reflects common alterations seen in human tumors, makes it an excellent model for evaluating the efficacy of potential drugs and novel therapeutic strategies. The use of D341 Med in these studies helps to bridge the gap between laboratory research and clinical application, supporting the development of targeted therapies that could offer improved outcomes for children affected by this devastating disease.

**Organism**

Human

**Tissue**

Brain, cerebellum

**Disease**

Medulloblastoma

**Synonyms**

D-341 Med, D-341 MED, D-341MED, D341\_Med, D341Med, D341MED, D341MD, D-341, D341, Med 341, H341

**Characteristics**

**Age**

3,5 years

**Gender**

Male

**Ethnicity**

European

**Morphology**

Lymphoblast

**Growth properties**

Suspension

**Regulatory Data**

**Citation**

D341Med (Cytion catalog number 305136)

**D341Med Cells | 305136****Biosafety level** 1**NCBI\_TaxID** 9606**CellosaurusAccession** CVCL\_0018**Biomolecular Data****Protein expression** Glutamine synthetase positive, neuron specific enolase positive, glial fibrillary acidic proteins negative, S100 (S-100) protein negative, neuroectodermal antigen positive, recognized by the UJ13A monoclonal antibody**Tumorigenic** Yes**Handling****Culture Medium** EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO<sub>3</sub>, w: EBSS (Cytion article number 820100a)**Supplements** Supplement the medium with 10% FBS and 1% NEAA**Doubling time** 37 hours**Subculturing** Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of  $1 \times 10^5$  cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.**Fluid renewal** 2 to 3 times per week**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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**Thawing and  
Culturing Cells**

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

**Incubation  
Atmosphere**

37°C, 5% CO<sub>2</sub>, humidified atmosphere.

**Shipping  
Conditions**

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

**Storage  
Conditions**

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

**Quality Control & Molecular Analysis**

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**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.