

## RT-112 Cells | 300324

## General information

<b>Description</b>	This cell line was established by Dr. Carol Rigby, St. Paul's Hospital, London as described by Benham et al. in 1976 from a human bladder carcinoma.
<b>Organism</b>	Human
<b>Tissue</b>	Bladder
<b>Disease</b>	Carcinoma
<b>Synonyms</b>	RT 112, RT112

## Characteristics

<b>Age</b>	Unspecified
<b>Gender</b>	Female
<b>Ethnicity</b>	Caucasian
<b>Morphology</b>	Epithelial-like
<b>Growth properties</b>	Adherent

## Regulatory Data

<b>Citation</b>	RT-112 (Cytion catalog number 300324)
<b>Biosafety level</b>	1
<b>NCBI_TaxID</b>	9606
<b>CellosaurusAccession</b>	CVCL_1670

## Biomolecular Data

<b>Protein expression</b>	p53 positive, Cytokeratine (4),5,(6), 7, 8, 13, 17, 18, 19, Desmoplakin
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**Isoenzymes** Yes, in nude mice

**MSI-status** Stable (MSS)

**Handling**

**Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)

**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

**Split ratio** A ratio of 1:4 to 1:8 is recommended

**Seeding density** Start culture at 2 to 3 x 10<sup>4</sup> cells/cm<sup>2</sup> and continue with a seeding density of 1 x 10<sup>4</sup> cells/cm<sup>2</sup>.

**Fluid renewal** 2 to 3 times per week

**Freeze medium** As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

**Quality Control & Molecular Analysis**

**Sterility** Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

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**STR profile**

**Amelogenin:** x,y  
**CSF1PO:** 10,11  
**D13S317:** 13,14  
**D16S539:** 11,13  
**D5S818:** 10,13  
**D7S820:** 12,11  
**TH01:** 7  
**TPOX:** 8,11  
**vWA:** 14,17  
**D3S1358:** 15  
**D21S11:** 27,3  
**D18S51:** 15  
**Penta E:** 12,16  
**Penta D:** 10,11  
**D8S1179:** 13,15  
**FGA:** 23

**HLA alleles**

**A\*:** '26:01:01  
**B\*:** '27:05:02  
**C\*:** '01:02:01  
**DRB1\*:** '01:01:01  
**DQA1\*:** '01:01:01  
**DQB1\*:** '05:01:01  
**DPB1\*:** '01:01:01  
**E:** '01:01:01