Product sheet

PC-12 Cells | 500311

General information

Description	PC-12 cells are a cell line derived from a pheochromocytoma of the rat adrenal medulla. These cells are of embryonic origin, grow adherently and resemble a mixture of neuroblastic and eosinophilic cells. PC-12 cells are catecholamine cells that synthesize, store and release norepinephrine and dopamine. They have a diameter of approximately 10-12 microns and are small, irregularly shaped cells. The PC12 cell line is a classical neuronal cell model due to its ability to acquire sympathetic neuron features when dealing with nerve growth factor (NGF). Studies on dopamine regulation have shown that PC12 cells synthesize, release, and reuptake dopamine and have been extensively characterized for neurosecretion and the presence of ion channels and neurotransmitter receptors. Moreover, the relative proportion of various subtypes of Ca channels changes during differentiation. The PC12 cell line is an established neuronal cell model that is particularly useful in studying cellular responses to nerve growth factors (NGF) and how these lead to the expression of differentiation-specific proteins and differentiation. When cultured in NGF, PC12 cells differentiate into sympathetic ganglion neurons morphologically and functionally. The differentiation results from the reversible induction of a neuronal phenotype by NGF. Collagen coating has been shown to be favourable to achieving neuronal characteristics in terms of length and density of neurites by NGF treatment. PC12 cells are tumorigenic and were derived from male New England Deaconess Hospital strain rats. The PC-12 cell line has 40 chromosomes, 38 autosomes, plus xY. Nerve growth factor (NGF) is expressed in PC12 cells, and exposure to NGF is one crucial regulator of cell differentiation. In conclusion, PC12 cells are a versatile and widely used model system in neurobiology due to their ability to acquire sympathetic neuron features when dealing with nerve growth factor (NGF). These cells have been extensively characterized for neurosecretion, ion channels, and neurotransmitter receptors. Their extreme versatility for pharmacological testing and use as an established model for studying the proliferation and differentiation of neuronal cells make them a valuable tool in neurobiology research.
Organism	Rat
Tissue	Adrenal gland
Disease	Pheochromocytoma
Synonyms	PC 12, PC12

Characteristics

Age	Unspecified
Gender	Male
Ethnicity	Japanese
Morphology	Polygonal

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Regulatory Data

Citation	PC-12 (Cytion catalog number 500311)
Biosafety level	1
NCBI_TaxID	10116
CellosaurusAccession	CVCL_S979

Biomolecular Data

Receptors expressed	Nerve growth factor (NGF)
Tumorigenic	Yes, in New England Deaconess Hospital strain rats
Products	Catecholamines, dopamine
Karyotype	40 chromosomes, 38 autosomes plus xY

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS
Subculturing	Suspension cells: Remove cells from substrate by pipetting with fresh medium. To obtain single cells, pass the suspension several times through a 22 gauge needle and dispense into new flasks. Growing on collagen: To remove adherent cells, use the following standard protocol. Remove medium and rinse the adherent cells using PBS without calcium and magnesium (3-5 ml PBS for T25, 5-10ml for T75 cell culture flasks). Add TrypleExpress (1-2ml per T25, 2.5ml per T75 cell culture flask), the cell sheet must be covered completely. Incubate at 37 degree Celsius for 10 minutes. Carefully resuspend the cells, the addition of medium is optional but not necessary, and dispense into new flasks which contain fresh medium.
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	2 to 3 times per week

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Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 48 hours.
Freeze medium	As a cryopreservation medium, we use 50% basal medium + 40% FBS + 10% DMSO, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Flask Coating	Collagen
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.