Product sheet

Capan-2 Cells | 300144

General information

Description	The Capan-2 cell line is a human pancreatic adenocarcinoma cell line first isolated from the pancreatic tumor tissue of a 56-year-old Caucasian male. It was derived from the metastatic site in the liver, indicating its origin from a secondary tumor which makes it particularly valuable for research on metastatic processes and pancreatic cancer biology. The cells exhibit epithelial morphology and have been utilized extensively to study pancreatic cancer, drug resistance, and tumor biology. Capan-2 cells are known to express a mutated form of the Kirsten rat sarcoma viral oncogene homolog (KRAS), a common mutation in pancreatic cancer, making them a robust model for studying KRAS-driven tumorigenesis. Additionally, they are characterized by the expression of tumor suppressor gene p53 mutations and have been observed to exhibit chromosomal instabilities, which are critical features relevant to cancer progression and treatment response. This cell line has been used in numerous studies, including those evaluating chemotherapeutic efficacy, exploring molecular pathways of cancer progression, and developing targeted therapy strategies.
Organism	Human
Tissue	Pancreas
Disease	Adenocarcinoma
Synonyms	CaPan-2, CAPAN-2, Capan 2, CAPAN 2, Capan2, CAPAN2

Characteristics

Age	56 years
Gender	Male
Ethnicity	Caucasian
Morphology	Polygonal
Growth properties	Adherent, colonies

Regulatory Data

Citation	Capan-2 (Cytion catalog number 300144)
Biosafety level	1

Capan-2 Cells | 300144

NCBI_TaxID	9606
CellosaurusAccession	CVCL_0026

Biomolecular Data

Protein expression	P53 negative
Antigen expression	Blood Type B, Rh+
Isoenzymes	Me-2, 2, PGM3, 2, PGM1, 1, ES-D, 1, AK-1, 1, G6PD, B, GLO-1, 2, Phenotype Frequency Product: 0.0004
Tumorigenic	Yes, in nude mice. Forms well differentiated adenocarcinoma consistent with pancreatic carcinoma
Products	Mucin (apomucin, MUC-1, MUC-2)
Ploidy status	Aneuploid
Mutational profile	Capan-2 cells carry a heterozygous Kras mutation in codon12: GGT>GTT

Handling

Culture Medium	McCoys 5a, w: 3.0 g/L Glucose, w: stable Glutamine, w: 2.0 mM Sodium pyruvate, w: 2.2 g/L NaHCO3 (Cytion article number 820200a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Doubling time	45 to 60 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Capan-2 Cells | 300144

Seeding density	1 x 10^4 cells/cm^2 will result in a confluent monolayer within 7 days.
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 48 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.

Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.