Product sheet

Hep-56.1C Cells | 400203

General information

Description	The Hep-56.1c hepatoma cell line is derived from a mouse liver tumor, specifically from the C57BL/6J mouse strain. This cell line is characterized by a notable mutation in the p53 gene, identified at different passages during in vitro propagation. Specifically, Hep-56.1c exhibits a C:G to G:C transversion at codon 132 of exon 5, resulting in an amino acid change from cysteine to tryptophan. This mutation was detected at passage number 17, suggesting a selective growth advantage conferred by the mutation, leading to its predominance in the cell population. The Hep-56.1c cell line displays a predominantly epithelial morphology, reflecting its hepatocytic origin. This is consistent with its intermediate filament protein profile, which includes the simple keratins K8 and K18, as well as vimentin and keratin K19 to varying degrees. The presence of these proteins confirms the hepatocytic nature of the cell line and its classification as a hepatoma line. Further analysis of Hep-56.1c using DNA fingerprinting did not reveal any major structural abnormalities, although some changes in the relative intensities of specific bands were observed with increasing passage numbers. This indicates genomic stability with some degree of variability over extended culture periods. The p53 mutation analysis and intermediate filament protein expression patterns together establish Hep-56.1c as a valuable model for studying hepatocellular carcinoma and the role of p53 mutations in liver tumorigenesis.
Organism	Mouse
Tissue	Liver
Disease	Hepatocellular carcinoma
Synonyms	HEP-56.1C, 56.1C, 56.1c

Characteristics

Breed/Subspecies	C57BL/6J
Age	Adult
Gender	Female
Morphology	Epithelial-like
Growth properties	Adherent

Regulatory Data

Hep-56.1C Cells | 400203

Citation	Hep-56.1C (Cytion catalog number 400203)
Biosafety level	1
NCBI_TaxID	10090
CellosaurusAccession	CVCL_5768

Biomolecular Data

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	Every 3 to 5 days
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Hep-56.1C Cells | 400203

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.