Product sheet

9L/lacZ Cells | 305208

General information

Description	The 9L/lacZ cell line is a well-characterized rat gliosarcoma cell line commonly used in neurobiological and oncological research. Originally derived from a nitrosourea-induced rat brain tumor, this line has been engineered to express the lacZ gene, which encodes the enzyme β-galactosidase. This modification facilitates the tracing and studying of tumor cells in vivo, particularly useful in experiments involving tumor progression and metastasis. The expression of lacZ allows for the easy identification of these cells using X-gal staining, which turns the cells blue when they express β-galactosidase. These cells exhibit aggressive tumor-forming capabilities when implanted in immunocompromised or syngeneic hosts, making them a robust model for studying brain cancer dynamics and testing therapeutic strategies against gliomas. Additionally, the 9L/lacZ cell line has been utilized in gene therapy trials, particularly in assessing the efficacy of suicide genes and other genetic interventions aimed at controlling tumor growth. This line is also pivotal in understanding the interactions between tumor cells and the host's immune system, thereby contributing insights into the complexities of tumor immunology.
Organism	Rat
Tissue	Brain
Disease	Rat malignant glioma
Synonyms	9L/LacZ

Characteristics

Breed/Subspecies	Fischer 344
Gender	Male
Morphology	Fibroblast
Growth properties	Adherent

Regulatory Data

Citation	9L/lacZ (Cytion catalog number 305208)
Biosafety level	1
NCBI_TaxID	10116

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CellosaurusAccession	CVCL_5656
GMO Status	GMO-S1: This rat glioma cell line (9L/lacZ) contains lacZ and Tn5-neo genes delivered via a replication-deficient BAG retroviral vector, enabling β-galactosidase expression and neomycin resistance. The modification is stable in 9L glioma cells. This classification applies only within Germany and may differ elsewhere

Biomolecular Data

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.