Product sheet

MDA-MB-157 Cells | 305280

General information

Description	The MDA-MB-157 cell line is derived from a human breast carcinoma, specifically from a pleural effusion of a metastatic breast cancer patient. This cell line is extensively used in breast cancer research, particularly for studying the biology of triple-negative breast cancer (TNBC), a subtype that lacks expression of estrogen receptor (ER), progesterone receptor (PR), and HER2/neu. MDA-MB-157 cells provide a valuable model for investigating the molecular mechanisms driving TNBC, as well as for testing potential therapeutic agents targeting this aggressive form of breast cancer. MDA-MB-157 cells exhibit an epithelial morphology and are characterized by their high metastatic potential. They express markers typical of basal-like breast cancer, including cytokeratins 5/6 and epidermal growth factor receptor (EGFR). Researchers utilize MDA-MB-157 cells to explore key signaling pathways involved in TNBC progression, such as the PI3K/Akt, MAPK, and Notch pathways. These cells are also employed in drug screening assays to evaluate the efficacy of chemotherapeutic agents, targeted therapies, and combination treatments. Additionally, MDA-MB-157 cells are used to study the mechanisms of drug resistance and to develop strategies to overcome it. The relevance of the MDA-MB-157 cell line in triple-negative breast cancer research underscores its importance in advancing our understanding of this challenging subtype of breast cancer and in developing more effective therapeutic approaches for TNBC patients.
Organism	Human
Tissue	Breast
Disease	Carcinoma
Metastatic site	Pleural effusion
Synonyms	MDA-MB157, MDAMB157, MDA-157, MDA157, MB 157, MB157, MD Anderson-Metastatic Breast-157

Characteristics

Age	44 years
Gender	Female
Ethnicity	African American
Morphology	Epithelial
Growth properties	Adherent

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Regulatory Data

Citation	MDA-MB-157 (Cytion catalog number 305280)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0618

Biomolecular Data

Surface antigens	Blood Type B, Rh -
Oncogenes	WNT7B +
Tumorigenic	Yes, in nude mice and in immunosuppressed BALB/c mice
Mutational profile	Mutation: MSH6, p.Pro42Ser (c.124C>T), heterozygous; Mutation: MSH6, p.Arg644Ser (c.1932G>C), heterozygous; Mutation: TP53, p.Pro87fs*53 (c.261_286del26) (p.Ala88Cysfs*52), homozygous

Handling

Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
Supplements	Supplement the medium with 20% FBS + Insulin (5 microgram/ml)
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week

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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Flask Coating	None
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.