

## SNU-878 Cells | 305285

## General information

## Description

The SNU-878 cell line is derived from a human hepatocellular carcinoma (HCC), which is a primary malignancy of the liver. This cell line is widely used in liver cancer research to study the molecular and cellular mechanisms underlying hepatocarcinogenesis, tumor progression, and therapeutic responses. Hepatocellular carcinoma is one of the most common and deadly forms of liver cancer, making cell lines like SNU-878 essential for advancing our understanding of the disease and developing effective treatments.

SNU-878 cells exhibit an epithelial morphology and express markers typical of liver cancer, such as alpha-fetoprotein (AFP) and hepatocyte-specific antigens. They harbor genetic and epigenetic alterations that are frequently observed in HCC, including mutations in key oncogenes and tumor suppressor genes. Researchers use SNU-878 cells to explore various signaling pathways involved in liver cancer, such as the Wnt/ $\beta$ -catenin, PI3K/Akt, and MAPK pathways. These cells are also employed in high-throughput drug screening assays and preclinical testing of chemotherapeutic agents, targeted therapies, and combination treatments. Additionally, SNU-878 cells are utilized to study mechanisms of drug resistance and to develop strategies to overcome it. The relevance of the SNU-878 cell line in hepatocellular carcinoma research highlights its importance in advancing our knowledge of liver cancer biology and in the development of new therapeutic approaches for patients with HCC.

## Organism

Human

## Tissue

Liver

## Disease

Adult hepatocellular carcinoma

## Synonyms

SNU878, NCI-SNU-878

## Characteristics

## Age

54 years

## Gender

Female

## Ethnicity

East Asian

## Morphology

Epithelial

## Growth properties

Adherent

## Regulatory Data

**SNU-878 Cells | 305285****Citation** SNU-878 (Cytion catalog number 305285)**Biosafety level** 1**NCBI\_TaxID** 9606**CellosaurusAccession** CVCL\_5102**Biomolecular Data****Mutational profile** Mutation: TP53, p.Ile251Asn (c.752T>A), homozygous; Mutation: TSC2, p.Ser1514Ter (c.4541C>G), homozygous**Handling****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)**Supplements** Supplement the medium with 10% heat-inactivated FBS, 25 mM HEPES**Dissociation Reagent** Accutase**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Fluid renewal** 2 to 3 times per week**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below  $-150^{\circ}\text{C}$  to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a  $37^{\circ}\text{C}$  water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at  $300 \times g$  for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Incubation Atmosphere

$37^{\circ}\text{C}$ , 5%  $\text{CO}_2$ , humidified atmosphere.

### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately  $-78^{\circ}\text{C}$  throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about  $-150$  to  $-196^{\circ}\text{C}$ . Storage at  $-80^{\circ}\text{C}$  is acceptable only as a short interim step before transfer to liquid nitrogen.

## Quality Control & Molecular Analysis

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**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.