

XXXXXXXXRF/6A | 305150

XXXXXXXXXX XXXXX

Description	XXXX XXXXXX XXXX-XXXX XXXXXX XXXXXXXX XXXXXXXXXXXX
Organism	XXXXX XXXXXX
Tissue	XXXXXXXXXX XXXXXXXX
Disease	Normal retinal choroidal endothelium (fetal; non-tumorigenic)
Metastatic site	Not applicable (normal fetal retinal choroidal endothelial cell line)
Applications	Ocular angiogenesis research; retinal and choroidal vascularization; anti-VEGF therapy evaluation (bevacizumab, ranibizumab); AMD and diabetic retinopathy modeling; tube formation assays; vascular permeability; NHP primate retinal endothelial model

XXXXXXXXXX

Age	XXXXXX
Gender	Sex unspecified
Ethnicity	Not applicable (non-human primate cell line; Macaca mulatta)
Morphology	XXXXXXXXXX
Cell type	Endothelial cells
Growth properties	XXXXXX

XXXXXXXXXXXX XXXXXXXXXXXXX

Citation	RF/6A (XXXXXXX XXXXXXX XXX XXXXXXXXXX 305150)
Biosafety level	1
NCBI_TaxID	9544
CellosaurusAccession	CVCL_4552

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GMO Status No genetic modification; wildtype rhesus macaque fetal retinal choroidal endothelial cell line

XXXXXXXXXXXX XXXXXXXXXXXX XXXXXXXXXXXX

Protein expression XXXXXXXX X XXXXXXXXXXXXXXXX

XXXXXXXXXXXX

Culture Medium EMEM (MEM Eagle) X 2 XXXXX XXXXX-XXXXXXXXXXXX X 2.2 X/XXXX NaHCO3 X EBSS (XXXX XXXXXXXX 820100a X XXXXXXXX)

Supplements XXXXXXX XXX XXXXXXX 10 X XXXXXXX XXXXX XXXXXXXX X1 X XXX XXXXXXXXXXXXXXX XXXXXXXX

Dissociation Reagent XXXXXXXX

Doubling time approx. 24 to 36 hours

Subculturing XX XXXXXXX XXXXXXX XXXXXXX XX XXXXXXX XXXXXXXX XXXXXXX XXXXXXXX PBS XXXXX XXXXXXX XXX XXXXXXXXXXXXXXX XXXXXXXXXXXXXXX XXXXXXXXXXXXXXX XXXXXXXXXXXXXXX

Split ratio 1:2 XXX 1:4

Seeding density 1 to 2 x 10⁴ cells/cm²

Fluid renewal 2 XXX 3 XXXXX XX XXXXXXXX

Post-Thaw Recovery After thawing, plate the cells at 5 x 10⁴ cells/cm² and allow at least 24 hours for adherence before the first medium change. Do not allow cultures to reach full confluency as contact inhibition may reduce endothelial phenotype.

Freeze medium XXXXXXX XXXXXXX XXXXXXXXXXXXXXX XXXXXXX XXX XXX XXXX (XXXX XX XXX FBS) + 10% DMSO XX XXX XXXXXXX XXX XXXXXXX XXXXXXX XXX XXXXXXX XXXXXXX

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Thawing and Culturing Cells

1. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
2. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
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4. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
5. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
6. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
7. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.
8. Thaw the vial in a water bath at 37°C. Do not shake the vial. Remove the vial from the water bath and centrifuge at 300 x g for 3 minutes. Discard the supernatant and resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. The cells should reach 70% confluency within 8-15 days. Pass the cells into a new T75 flask when they reach 70-80% confluency.

Incubation Atmosphere

37°C, 5% CO2

Flask Coating

None

Freezing Procedure

Freeze cells in complete medium supplemented with 10% FBS and 10% DMSO at -80°C.

Shipping Conditions

Store at -80°C. Ship on dry ice.

Storage Conditions

Store at -150 to -196°C.

HLA

Sterility

The cells are tested for mycoplasma contamination using PCR. The cells are free of mycoplasma contamination.