

### U2OS-CRISPR-TPR-SNAP | 300667

#### U2OS-CRISPR-TPR-SNAP

**Description** U2OS-CRISPR-TPR-SNAP is a cell line derived from U2OS cells expressing a CRISPR-Cas9 system targeting the TPR gene. The cells are stably transfected with a CRISPR-Cas9 system targeting the TPR gene. The CRISPR-Cas9 system is used to generate a TPR knockout cell line. The cells are then screened for TPR knockout using Western blot analysis. The TPR knockout cell line is then used for the study of TPR function in cell cycle regulation and DNA damage response.

**Organism** Homo sapiens

**Tissue** U2OS

**Disease** None

**Metastatic site** None

**Applications** Western blot analysis, RT-PCR, qPCR, CRISPR-Cas9 screening (mRNA), TPR knockout, cell cycle regulation, DNA damage response

#### U2OS-CRISPR-TPR-SNAP

**Age** 15 days

**Gender** Male

**Ethnicity** Caucasian

**Morphology** Adherent, fibroblastic

**Cell type** U2OS (CRISPR-Cas9 targeting TPR)

**Growth properties** None

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**Citation** U2OS-CRISPR-TPR-SNAP (CRISPR-Cas9 targeting TPR) | 300667

**Biosafety level** 1

**NCBI\_TaxID** 9606

Product sheet

**U2OS-CRISPR-TPR-SNAP | 300667**

**CellosaurusAccession**   **U2OS-CRISPR-TPR-SNAP** (U2OS CRISPR U2OS CVCL\_0042)

**Depositor**   **EMBL** (EMBL)

**GMO Status**   GMO-S1: U2OS-CRISPR-TPR-SNAP (U2OS-CRISPR-TPR-SNAP)

**Protein expression**   TPR SNAP-tag

**Culture Medium**   5% FBS, 3.0%  $\beta$ -MEM, 2.0%  $\alpha$ -MEM, 2.2% NaHCO<sub>3</sub>

**Supplements**   10% FBS, 3.0%  $\beta$ -MEM, 2.0%  $\alpha$ -MEM, 2.2% NaHCO<sub>3</sub>

**Dissociation Reagent**   Trypsin

**Doubling time**   24-36 hours

**Subculturing**   1:3 split ratio into PBS

**Split ratio**   1:3

**Seeding density**    $1 \times 10^4$  cells/cm<sup>2</sup>

**Fluid renewal**   2-3 times per week

**Freeze medium**   10% FBS + 10% DMSO

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### Thawing and Culturing Cells

1. Thaw the cells in a 37°C water bath. Transfer the cells to a pre-warmed medium.
2. Seed the cells into a 24-well plate at a density of 100,000 cells per well in 1 ml of medium.
3. Incubate the cells for 24 hours at 37°C in 5% CO<sub>2</sub>.
4. Remove the medium and replace it with fresh medium. The cells should reach 70% confluency.
5. Seed the cells into a 96-well plate at a density of 10,000 cells per well in 100 µl of medium.
6. Incubate the cells for 24 hours at 37°C in 5% CO<sub>2</sub>.
7. Harvest the cells and perform a Western blot analysis.
8. Repeat the experiment with different CRISPR-Cas9 constructs.

**Incubation Atmosphere** 37 °C, 5% CO<sub>2</sub>

**Flask Coating** No coating

**Freezing Procedure** Harvest cells and freeze in 10% FBS, 90% FCS at -80°C.

**Shipping Conditions** Dry ice, -78°C

**Storage Conditions** -150 to -196 °C

### Genotype / HLA

**Sterility** The cells are free of mycoplasma contamination. PCR screening for mycoplasma is recommended.