

Sp2/0-14 | 400481

General Information

Culture Medium DMEM 4.5 g/l, Glucose 4 g/l, L-Glutamine 3.7 g/l, NaHCO₃ 1.0 g/l, Penicillin 100 IU/ml, Streptomycin 100 µg/ml, Fungizone 0.025 µg/ml (all antibiotics in 100% ethanol)

Supplements 10% FBS

Subculturing Cells are detached using trypsin-EDTA and resuspended in PBS. Cells are then seeded into new flasks.

Seeding density 1 × 10⁵ cells per flask

Fluid renewal 2-3 times per week

Freeze medium DMEM + 10% FBS + 10% DMSO

- Thawing and Culturing Cells**
1. Thaw the vial in a 37°C water bath.
 2. Dilute the cells into 10 ml of DMEM + 10% FBS.
 3. Seed the cells into a 25 cm² flask.
 4. Allow the cells to attach for 24 hours.
 5. Remove the medium and replace with fresh DMEM + 10% FBS.
 6. After 24 hours, the cells should be 70% confluent.
 7. Pass the cells into a new flask.
 8. Repeat the process.

Incubation Atmosphere 37°C, 5% CO₂

Flask Coating None

