Product sheet

UM-SCC-112 | 305720

[redacted]

Description	UM-SCC-112 [redacted] (HNSCC) [redacted] (HNSCC). [redacted] UM-SCC-112 [redacted] (HNSCC) [redacted]
Organism	[redacted]
Tissue	[redacted]
Disease	[redacted] (HNSCC)
Applications	[redacted] (HNSCC) [redacted]

[redacted]

Age	[redacted]
Gender	[redacted]
Ethnicity	[redacted]
Morphology	[redacted]
Cell type	[redacted]
Growth properties	[redacted]

[redacted]

Citation	UM-SCC-112 ([redacted] Cytion: 305720)
Biosafety level	1
NCBI_TaxID	9606

[redacted]

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Culture Medium	DMEM + 4.5 g/l Glucose + 4 mM Glutamine + 3.7 g/l NaHCO3 + 1.0 mM Sodium Pyruvate (820
Supplements	Add 10% Fetal Bovine Serum (FBS) + 1% Non-Essential Amino Acids (NEAA)
Dissociation Reagent	Accutase
Doubling time	Approx. 24 - 48 hours
Subculturing	Wash cells with PBS and incubate with Accutase at 37 °C for 8–10
Split ratio	1:3 - 1:5
Seeding density	1 - 3 × 10^4 cells/cm^2
Fluid renewal	Every 2 - 3 days
Freeze medium	Cell culture medium + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a water bath at 37 °C.
2. Transfer cells into 9 - 10 ml pre-warmed medium and centrifuge at 150 x g for 5 minutes.
3. Discard supernatant and resuspend cells in fresh medium. Transfer cells into culture flask and incubate at 37 °C.
4. Change medium after 24 hours to remove residual DMSO and non-adherent cells. Continue culturing cells in fresh medium.
5. Cells should be passaged when they reach 70 - 90% confluency. Remove medium and wash cells with PBS. Add 1 - 3 ml Accutase and incubate at 37 °C for 5 - 15 minutes.
6. Add 8 - 10 ml medium and centrifuge cells at 200 x g for 5 minutes. Discard supernatant and resuspend cells in fresh medium. Transfer cells into culture flasks at a split ratio of 1:3 - 1:5.
7. Change medium every 2 - 3 days.

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Incubation Atmosphere	37
Shipping Conditions	-78
Storage Conditions	-150 -196