

Sf9 | 604328

Virus susceptibility

Highly susceptible to Measles virus (MeV), Rubella virus (RV), and Morbilliviruses (MNPV) [1].

Culture Medium

DMEM (high glucose, no serum)

Supplements

10% FBS, 2% FCS, 1% BSA, 1% HSA, 1% LPS, 1% Penicillin, 1% Streptomycin, 1% Nystatin

Dissociation Reagent

Trypsin

Subculturing

Subculture into fresh medium every 15-20 days. Passages should be performed at 70-80% confluency.

Seeding density

1×10^4 cells/cm². Seed cells at 26-30% confluency. Seed cells into fresh medium every 15-20 days.

Fluid renewal

2-3 times per week

Freeze medium

DMEM (high glucose, no serum) + 10% DMSO + 10% FBS + 10% FCS + 10% BSA + 10% HSA + 10% LPS + 10% Penicillin + 10% Streptomycin + 10% Nystatin

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Thawing and Culturing Cells

1. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
2. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
3. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
4. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
5. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
6. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
7. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.
8. Thaw the vial rapidly in a 37 °C water bath. Do not vortex. Remove the vial from the water bath and centrifuge at 300 × g for 3 min. Transfer the cells to a new vial and centrifuge at 300 × g for 3 min. Resuspend the cells in 10 ml of complete medium. Seed the cells into a T75 flask containing 10 ml of complete medium. Incubate the cells for 15 days. Harvest the cells at day 8.

Incubation Atmosphere 27°C, 0% CO₂, humidified atmosphere.

Shipping Conditions Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

/ / HLA

Sterility [Detailed sterility testing information]