

MDA-MB-435S | 300277

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_0622

GMO Status No genetic modification; problematic line — parental MDA-MB-435 identified as M14 melanoma derivative; use with appropriate caution and cite genetic identity

XXXXXXXXXX XXXXXXXXXXXX XXXXXXXXXXXX

XXXXXXXXXX

Culture Medium DMEM: DMEM:Ham's F12 (1:1) 3.1 µg/ml transferrin 2.5 µg/ml selenium 15 µg/ml insulin (15 µg/ml transferrin 2.5 µg/ml selenium)

Supplements 5% FBS

Dissociation Reagent Trypsin

Subculturing 1:2 to 1:5 in DMEM:DMEM:Ham's F12 (1:1) 3.1 µg/ml transferrin 2.5 µg/ml selenium 15 µg/ml insulin (15 µg/ml transferrin 2.5 µg/ml selenium) + 5% FBS

Split ratio 1 to 5

Seeding density 1 to 3 × 10⁴ cells/cm²

Fluid renewal 2 to 3 times per week

Freeze medium DMEM:DMEM:Ham's F12 (1:1) 3.1 µg/ml transferrin 2.5 µg/ml selenium 15 µg/ml insulin (15 µg/ml transferrin 2.5 µg/ml selenium) + 5% FBS + 10% DMSO

MDA-MB-435S | 300277

Thawing and Culturing Cells

1. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
2. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
3. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
4. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
5. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
6. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
7. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.
8. Thaw the vial in a water bath at 37°C. Transfer the cells to a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 mL of complete medium. Seed the cells into a T75 flask and incubate at 37°C in 5% CO2. Once the cells reach 70% confluency, passage them into a new T75 flask.

Incubation Atmosphere 37°C, 5% CO2

Flask Coating None

Freezing Procedure Harvest cells into a 15 mL centrifuge tube and centrifuge at 300 x g for 3 minutes. Resuspend the cell pellet in 1 mL of freezing medium. Transfer to a cryovial and freeze in a programmable freezer.

Shipping Conditions Ship at -78°C

Storage Conditions Store at -150°C to -196°C

MDA-MB-435S / HLA

Sterility The cells are free of mycoplasmas and other contaminants. PCR testing confirmed the absence of mycoplasmas.